Curriculum Vitae

Gary J. Pielak

Department of Chemistry University of North Carolina at Chapel Hill Chapel Hill, NC 27599-3290 Phone: 919-962-4495 Email: gary_pielak@unc.edu Home page: www.chem.unc.edu/people/faculty/pielak/group/

Research Interests:

Protein chemistry and biophysics, especially in living cells.

Employment:

July 2015-

Kenan Distinguished Professor of Chemistry

August 2013-2018

Vice Chair of Facilities in Chemistry

July 2013-July 2015

Glen H. Elder, Jr. Distinguished Term Professor of Research & Undergraduate Education

January 2013-December 2014

Program Director, Molecular Biophysics Cluster, Division of Molecular and Cellular Sciences, Directorate for Biological Sciences, the National Science Foundation, Arlington, NC

January 2000-

Professor of Chemistry, UNC Professor of Biochemistry & Biophysics, UNC Member, UNC Lineberger Comprehensive Cancer Center

July 2000 - June 2005 & July 2007 - June 2008

Vice Chair of Undergraduate Studies in Chemistry

January 2000-June 2006

Faculty Director, UNC Macromolecular Interactions Facility

July 1998 - July 1999

Sabbatical Visitor: Professor Christopher M. Dobson, F.R.S. Oxford Centre for Molecular Sciences University of Oxford, England

January 1995-December 1999

Associate Professor of Chemistry, UNC

January 1989-June 2016

Co-director UNC Biomolecular NMR Facility

January 1989 - December 1994

Assistant Professor, UNC

Member UNC Integrative Program for Biological & Genome Sciences (previously Program in Molecular Biology and Biotechnology)

June 1986 - January 1989

N.I.H Postdoctoral Fellow: R.J.P. Williams, F.R.S., M.B.E, deceased Inorganic Chemistry Laboratory, University of Oxford, England

March 1983 - June 1986

N.I.H., Postdoctoral Fellow: Professor M. Smith, F.R.S., deceased Department of Biochemistry University of British Columbia Vancouver, B.C. Canada

Education:

January 1983

Ph.D. in Biochemistry Laboratory of Professor J. Ivan Legg, deceased Department of Chemistry Washington State University, Pullman, Washington Dissertation: Characterization of Arsanilazo & Sulfanilazo Proteins

June 1977

B.A. in Chemistry, *Magna cum Laude* Bradley University, Peoria, Illinois

Teaching/Mentoring:

Past Postdocs and Their Current Employer

Dr. Guifang Wang (Pennsylvania State University) Dr. Conggang Li (Chinese Academy of Sciences, Wuhan) Dr. Austin Smith (Jaguar Gene Therapy) Dr. Thomas Boothby (U. Wyoming)

Current Postdocs

None

Current Graduate Students

Mr. Oskar Hutcheson Ms. Julia Brom Ms. Jordyn Markle

Past Graduate Students and Their Current Employer

Dr. Claire Stewart (Stanford) Dr. I-Te Chu (Harvard) Dr. Jonathan (Jack) Eicher Dr. Joseph 'Joey' Thole (NIH) Ms. Sasiprapa Jeab Sasiprapa (visitor from Thailand) Dr. Candice Crilly Dr. Shannon Speer (Pfizer) Dr. Samantha (Sam) Stadmiller (Lindy Biosciences) Dr. Samantha (Pixie) Piszkiewicz (UC Berkelev) Dr. Alex Guseman (Pitt) Dr. Annelise Gorensek (Colorado College) Dr. Rachel Cohen (SprinWorks Therapeutics) Dr. Austin Smith (Jaguar Gene Therapy) Dr. Michael Senske (visitor from Germany, FC Bayern München) Ms. Torii Sutherland (US Coast Guard) Dr. William Monteith (Alector) Dr. Mohona Sarkar (KBI) Dr. Jillian Tyrrell (Biocare Medical) Dr. Yaqiang Wang (Arrakis Therapeutics) Ms. Laura Benton (Holy Orders) Dr. Imola-Gabriela Zigoneanu (Biomedical Engineering, UNC) Dr. Alex Schlesinger (AgBiome) Dr. Andrew Miklos (NIH) Mr. Christopher Barnes, M.A. (Stanford)

Past Graduate Students and Their Current Employer, cont'd

Dr. Rebecca Ruf Mr. Matthew Hrabak, M.S. (Naval Surface Warfare Center) Dr. Kristin Slade (Hobart and William Smith Colleges) Dr. Lisa Charlton (ERT) Dr. Brian McNulty (Athenix) Dr. Julie Bryant (Merck) Dr. Alina Olteanu (Private practice, Houston) Dr. Dana Albon (Moses Cone Mem. Hosp.) Dr. Fang Yi (Centocor) Dr. Chetan Patel (Lilly) Dr. Artemiza Morar (GlaxoSmithKline) Mr. Xuming Wang, M.A. (Intel) Dr. Gresham Weatherly (AbbVie) Dr. Paula Davis-Searles (Diosynth) Dr. Jennifer Waldner Ms. Devon Allen, M.S. (Diosvnth) Dr. David Cohen (Advanced Liquid Logic) Dr. Aleister Saunders (Drexel) Dr. Jennifer Marmorino Dr. Donald Dovle (Georgia Tech.) Dr. James Beasley, (Venenum) Dr. Lixin Chen (New Engl. Biolabs) Dr. Zoev Fredericks (Amgen) Dr. Douglas S. Auld (Novartis) Dr. Stephen F. Betz (Crinetics) Dr. Sharon Hilgen-Willis (Integral Molecular) Ms. Xuhong Wang, M.A. (Synermore Biologics)

Past STEM Teacher-researcher Fellows

Ms. Candice Jackson, Thomasville High School (2011) Mr. Oryan Lowry, South Robeson High School (2012)

Current Undergraduate Students

Ms. Ruta Petrikis

Mr. Thomas Redvanly

Mr. Owen Young

Mr. Joshua Bourque

Past Undergraduates and Where They Went Next

Mr. Hudson Malsch (UNC)

Ms. Ashlee Propst (UNC)

Ms. Penelope Mewborn (UNC Nutrition)

Ms. Shikun 'Rinco' Wang (Yale)

Ms. Sophia Hazlett (University College London)

Mr. Francis Lauzier (Wayne State)

Mr. Octavio Origel (Northwestern)

Mr. Owen Warmuth (Wisconsin)

Ms. Shreya Nakkala (UNC)

Mr. Jhoan Aguilar (Postbac UNC)

Mr. Gustavo Panduro (UNC)

Mr. Gerardo Perez (MIT)

Mr. Cody Weyhrich (Virginia Tech)

Mr. Aakash Mehta (NIEHS)

Mr. Anthony Arrington (UNC Pembroke)

Past Undergraduates and Where They Went Next, cont'd

Mr. Kenny Nguyen (U Wyoming)

Mr. Luis Acosta (Actuarial School)

Mr. Thomas Lanier (UNC)

Mr. Larry Zhou (NIH)

Mr. Eduardo Guizan (Pharmacy School, UNC)

Mr. Emilio Guzman (Med. School, UNC)

Mr. Joe Lu (Actuary School)

Ms. Beth Willard (Disney World)

Mr. Vishavpreet 'Ricky' Singh (Med. School, UNC)

Mr. Alexander Krois (Grad. School, Scripps)

Ms. Yuri Yang (Technician, U. Toronto)

Ms. Amanda Rosett (SURE, back to Susquehanna U.)

Ms. Kristen Black (Colegio Bilingüe New Horizons, Dominican Republic)

Mr. Emmanuel Chan (Technician, UNC)

Ms. Heidi Scronce (Technician, Duke)

Ms. Niama Sharaf (Grad. School, Pitt)

Mr. Christopher Barnes (Grad. School, Chemistry, UNC)

Mr. Evan Lutz (Med. School, ECU)

Ms. Sandy An (MD/PhD program, Wake Forest, U.)

Mr. Hao Wu

Ms. Hayley Fischer (Med. School, ECU)

Ms. Michelle Mian (Dental School, Harvard)

Ms. Essraa Bayoumi

Mr. Michael Minder (Med. School, Duke)

Mr. Chris Kragel (Med. School, ECU)

Mr. Devin Barrett (Grad. School, Chemistry, UNC)

Mr. Joseph Batchelor (University of California, Berkeley)

Mr. Matthew Dedmon (Grad. School, University of Cambridge, UK)

Mr. Scott Kennedy (Grad. School, UNC)

Ms. Maria Lind (Grad. School, UGA)

Mr. Ikey Kakouras (Duke)

Ms. Kimberly Clay (Med. School, UNC)

Ms. Amret Thompson (Med. School, Wake Forest)

Mr. Daniel Hostetter (Grad. School, Stanford)

Ms. Melisa Lehti (Grad. School, Botany, Wisconsin)

Mr. Phil Hardwidge (Grad. School, Immunology, Mayo Clinic)

Mr. Sherif Ghobrial (Grad. School, Env. Sci. & Eng., UNC)

Mr. Chetan Patel (Grad. School, Chemistry, UNC)

Ms. Sonja Trojak (Med. School, UNC)

Mr. Bryan Fine (Med. School, U South Florida)

Ms. Xecerla Littles (Med. School, Tulane)

Ms. Shelly Finger (Vet. School, Texas A&M)

Mr. Luiz Alcazar-Roman (Grad. School, Chemistry)

Mr. Harvey Chui (Med. School, UNC)

Ms. Kara Bortone (Grad. School, Chemistry, U. Texas)

Mr. Sudip Parikh (Grad. School, Biochem., Scripps)

Mr. Richard Bruick (Grad. School, Biochem., Scripps)

Ms. Andrea Lee (Grad. School, Biochemistry, Scripps)

Mr. Mark Dransfield (Med. School, UNC)

Ms. Jennifer Fencl (Grad. School, Chemistry, UNC)

Ms. Tori Williams (Grad. School, Yale)

Ms. Malika Rauf (back to North Chapel Hill High)

Ms. Melanie Wiley (U. Maryland, then MD/PhD program, U. South Carolina)

Ms. Ashlee Propst (NC State)

Past High School Students and Where They Went Next

Ms. Hanna Qu (Research Triangle High) Ashlee Propst (NC State)

Courses:

UNC

Advances in Macromolecular Structure Macromolecular Structure and Metabolism Macromolecular Interactions Practical Protein NMR First Semester General Chemistry First Year Seminar: You don't have to be a rocket scientist. General Biochemistry Protein Chemistry Enzyme Mechanisms Molecular Biology Laboratory Practical Oligonucleotide-Directed Mutagenesis

Oxford

Biophysics Tutor. New College, 1988

Cold Spring Harbor

Advanced Cloning Course, 1984, 1987 Advanced Techniques in Molecular Biology

University of British Columbia

Site-specific Mutagenesis Directed by Oligodeoxyribonucleotides, 1985

Service:

Current Committee Assignments in Chemistry

Personnel Committee Teaching Assistant Professor Search Committee Staff Awards/Recognition Approximately 10 Ph.D. Committees Approximately 5 Undergraduate Honors Committees

Past Committee Assignments in Biochemistry & Biophysics

Biophysics Search Committees; Campbell, Lee, and Kuhlman Biomolecular NMR core director search, 2018 Assistant Professor search, UNC Biochemistry & Biophysics, 2019

Past Committee Assignments in Chemistry

Personnel Committee Teaching Assistant Professor Search Committee 2019 Executive Committee, 2015-2020 CHEM 550L Efficacy Committee, 2019-2019 X-Ray Core Director Search, 2018 NMR Core Search, 2018, 2019 Mass Spec Core Search, 2018 Vice Chair of Chemical Research Instrumentation Teaching & Core Laboratories, 2014-2018 Lecturer Search, 2016-2017 Mass Spectroscopy Core Director Search, 2014 Post Tenure Review Committee, 2011-2012, 2020-Graduate Studies Committee, 2010-2011 Graduate Recruiting Committee, 2010-2011

Past Committee Assignments in Chemistry, continued

Strategic Planning Committee, 2009 - 2010 2010 Departmental Program Review Committee NMR Committee Undergraduate Studies Committee, 1992-2008, 2012-2013 Inorganic Search Committee, 2009 Vice Chair of Undergraduate Studies, 2000-2005, 2007- 2008 Chair Selection Committee, 2007 Ad hoc member Parking Committee Search Committees; Forbes, Thorp, Erie, Morken, and Weeks Genomics Search Committee, 2001 Several Promotion/Tenure Committees

Current/Past University Service

University Teaching Awards Committee, 2018, 2019, 2020, 2023 University Distinguished Professorship Committee, 2017 Distinguished Dissertation Faculty Review, 2014, 2015 COI Monitoring Committee (Redinbo), 2014 Cross-listed Courses Task Force, 2012 Independent Studies Task Force, 2011 Chair, Curriculum Review Committee, Miscellaneous Subcommittee, 2010 University Research Day Judge, 2010 Chair, Admissions Committee, Biological & Biomedical Sciences Program, 2008 Administrative Boards of the General College, 2003-2013 Mock interviews for the Gates-Cambridge and Churchill Fellowships through the Office of Distinguished Scholarships, 2007 Reviewer, Smallwood Undergraduate Summer Research Grants, 2006 Summer Undergraduate Research Fellowship Committee, 2007-2010, 2015, ...2018, 2023 Mock interviews for the Gates-Cambridge and Churchill Fellowships through the Office of Distinguished Scholarships, 2007 Undergraduate Orientation (CTOPS) Professor's Perspective sessions, 2006, 2007, 2008, 2015, 2016, 2017 Financial Exigency and Program Change Committee, 2006-2008 Reviewer, Postdoctoral Awards for Research Excellence, Office of Postdoctoral Services, 2005 Division of Natural Sciences Curriculum Committee General Education Implementation Committee for the New Undergraduate Curriculum, 2002 - 2003 UNC Curriculum Review, Committee N Faculty Council **Rhodes Scholarship Mock Interview Committee Churchill Scholarship Selection Committee** Admissions Committee, Program in Molecular & Cellular Biophysics Advisory Committee, Curriculum in Applied Sciences **Biomolecular NMR Facility Committee** Macromolecular Interactions Facility Committee Summer Undergraduate Research Program Selection Committee Graduate Student Committees: **Environmental Sciences** Engineering, Biochemistry & Biophysics, Immunology/Microbiology Cell & Developmental Biology Presenter, Project Uplift

Presenter, NC Renaissance Program

Regional/National/International Service

2019 Judge Annual Biomedical Research Conference for Minority Students (ABRCMS)

2016 Outside Honors Examiner, Hobart and William Smith Colleges 2016 Chair, Biophysics *in vivo* subgroup of the Biophysical Society 2015 Chair-elect, Biophysics *in vivo* subgroup of the Biophysical Society

Regional/National/International Service, continued

2011-2015, Member at Large, Biophysics *in vivo* subgroup of the Biophysical Society

2013-2014 Program Director, Molecular Biophysics Cluster, Division of Molecular and Cellular Sciences, Directorate for Biological Sciences, the National Science Foundation

Faculty Search Committee Biochemistry, Washington State University, 2003 One or more tenure/promotion letters per year

Ph.D. Committees

Duke, Georgia Tech., University of Barcelona, Yale Thesis examiner, Indian Institute of Technology, Indore

Manuscript Referee

ACS Chemical Biology Angewandte Chemie Archives of Biochemistry and Biophysics Biochemistry Biochimica et Biophysica Acta Accounts of Chemical Research **Biochimie Biomacromolecules Biophysical Chemistry Biophysical Journal** Biopolymers BioTechniques Biotechnology **BMC** Biology ChemBioChem **Chemical Neuroscience Chemical Physics Letters Coordination Chemistry Reviews** Crystal Growth & Design **FEBS Letters** Folding and Design Frontiers of Molecular Biosciences Inorganica Chimica Acta Inorganic Biochemistry International Journal of Biological Macromolecules Journal of the American Chemical Society Journal of Biological Chemistry Journal of Biological Inorganic Chemistry Journal of Biomolecular NMR Journal of Chromatography Journal of Inorganic Biochemistry Journal of Chromatography

Manuscript Referee, continued

Journal of Magnetic Resonance Journal of Molecular Biology Journal of Physical Chemistry Journal of Physical Chemistry Letters Journal of Proteome Research Macromolecules Magnetic Resonance Letters **Molecular Pharmaceutics** Nature Nature Methods Nature Structural Biology Nucleic Acids Research **Physical Chemistry Chemical Physics Protein Science** Proteins: Structure, Function, Genetics/Bioinformatics Proceedings of the National Academy of Sciences, U.S.A. Scientific Reports Softmatter

Editorial Duties

1998-1999	Paper Alerts contributor, Current Opinions in Structural Biology
2011-	Editorial Advisor, BMC Biophysics
2017-	Editorial Advisory Board. Protein Science
2021-	Editorial Board, Magnetic Resonance Letters

Proposal Review

Panels

15
Graduate Women in Science Scholarships
Internal Review for Lockheed Martin 2010 University Research Initiative
University Cancer Research Fund, 2009
NIH New Innovators Award, 2008, 2009
NSF Molecular Biochemistry Review Panel. Many times.
NIH Special Emphasis Panel to review proposals in response to RFQ NIH
ES2007006, entitled "Scientific Research Analysis," 2007
NIH NIH-NIDDK, Kidney, Urologic and Hematologic Diseases D
Sub Committee, 2007
Gordon Research Foundation, 2005
NIH Physical Biochemistry Study Section, Ad hoc, 1996
NIH Metallobiochemistry Study Section, Ad hoc, 2001
NIH Special Emphasis Panel: Technology Development for Biomedical
Applications, 2001
NIH Macromolecular Structure & Function A Study Section, Ad hoc, 2011

Ad Hoc, Mail/Email Reviews

Stanford Synchrotron Radiation Lightsource Czech Science Foundation Danish Council for Independent Research Experimental Program to Stimulate Competitive Research, French National Research Agency (ANR) Israel Science Foundation National Science Centre Poland Netherlands Organisation for Scientific Research NSERC Canada

Ad Hoc, Mail/Email Reviews, continued

Petroleum Research Fund Research Corporation Switzerland: ETH Zurich Research Commission UK: BBSRC, MRC, Wellcome U.S.: AAAS Research Competitiveness Program, Cottrell NSF, Nebraska Wellcome trust/DBT India Alliance

Meetings Organized/Convened

ACS Spring National Meeting, Formulating Biologics: from Laboratory to Market, March 17-23, 2024 8th International Symposium on the Higher Order Structure of Protein Therapeutics (HOS), San Mateo, 2019 Program Committee, Protein Society Symposium, Barcelona, 2015 Program Committee Chair, 26th Annual Protein Society Symposium, 2012 Biophysics Society Subgroup, Biopolymers in vivo, 2012 Chemistry Spectrum: recruiting high school students interested in science to UNC. 2008 Co-chair Proteins Gordon Research Conference, 2007 Vice Co-chair Proteins Gordon Research Conference, 2005 Triangle Biophysics Symposium, 1998 Glaxo-Wellcome UNC Symposium, 1989, 1998 Protein Structure Minisymposium, 1993 Southeastern Magnetic Resonance Conference, 1993 Second Carolina Conference on Protein Engineering, 1989

Research: Current Grants

Mechanism of protein protection by desiccation-tolerance molecules Source: NSF, CHE, CLP 2203505 Total award amount: \$495,000 Total period covered: 09/01/2022-08/31/2025

Recent Grants

Macromolecular Crowding and Protein Stability In Vitro and in Cells Source: NSF, MCB 1909664 Total award amount: \$968,151 Total period covered: 08/01/19-07/31/23

Impact of cosolutes on protein folding Source: United States - Israel Binational Science Foundation Total award amount: \$216,000 Total period covered: 08/01/18-07/31/22

Protein stabilizers from tardigrades Source: National Institutes of Health 1 R01 GM127291-01A1 Total award amount: \$1,172,000 Total period covered: 12/01/2018 - 11/30/2021

MRI: Acquisition of a Mass Spectrometer (Co-PI) Source: NSF 1726291 Total award amount: \$1,157,551 Total period covered: 08/15/2017 - 07/31/2020

MRI: Purchase of a 600 MHz spectrometer for high-sensitivity NMR Source: NSF CHE 1828183 Total award amount: \$444750 Total period covered: 08/01/18 - 07/31/2020

Protein stabilizers from tardigrades Integrative Program for Biological and Genome Sciences Total award amount: \$30,000 Total period covered: 09/01/2018-08/31/2019

Tardigrade proteins as Novel Pharmaceutical Excipients Source: North Carolina Biotechnology Center Total award amount: \$75,000 Total period covered: 07/01/18-06/30/19

Macromolecular Crowding and Protein Stability In Vitro and in Cells Source: NSF MCB 1410854 Total award amount: \$ 990,000 Total period covered: 09/01/14-08/31/19

Encapsulation and Protein Stability Source: NSF CHE 1607359 Total award amount: \$ 353927 Total period covered: 07/01/2016-06/30/2018

Intergovernmental Mobility Award

Source: NSF 1410854 Total award amount: \$171,167 Total period covered: 12/31/12 – 06/31/13

Macromolecular Crowding and Protein Stability In Vitro and in Cells Source: NSF MCB 1051819 Total award amount: \$ 792,597.00 Total period covered: 02/01/11-01/31/14

E.T.S. Walton Visitor Award: Protein Chemistry in Living Cells Source Science Foundation of Ireland Total award amount: \$57,484 Total period covered: 01/01/12-12/30/12

In-Cell NMR of Disease-Related Proteins NIH Pioneer Award 5DP1OD783 Total award amount: \$3,750,000 Total period covered: 10/01/2006 – 09/31/2011

Protein Biophysics in Cells, Source: NSF MCB 0516547 Total award amount: \$592,931 Total period covered: 03/01/2006 – 02/28/2009

Electron Transfer Proteins

Source: NIH R01GM020488 (Francis Millett, PI) Total award amount: \$87,300 (to my laboratory) Total period covered: 08/01/03 - 07/31/08This was a subcontract to Professor Francis Millet's NIH grant. Prof. Millett is at the University of Arkansas. Our laboratories have collaborated on protein electron transfer for over 10 years. The funds support our work to produce cytochrome c variant proteins.

Perturbation Calorimetry & Protein Surface Area Source: PRF 42748-AC4 Total award amount: \$80,000 Total period covered: 05/01/05-08/31/07

Protein Biophysics in Cells Source: NSF MCB 0212939 Total award amount: \$446,735 Total period covered: 09/01/02 – 08/31/05

Patterned Library Analysis Source: NIH R01GM058665 (Marshall Edgell, PI) Total award amount: \$ 873,000 Total period covered: 07/01/00 – 06/30/04 co-PI with Marshall Edgell on this grant.

Free Radicals, Proteins Aggregates & Parkinson's Disease Source: NIH R21 ES 10774 Total award amount: \$290,000 Total period covered: 10/01/00 – 9/30/02

Cytochrome c & Apoptosis Source: NSF MCB0109366 Total award amount: \$145,000 Total period covered: 9/01/01-8/31/02

Expansion of the UNC Macromolecular Interactions Facility Source: North Carolina Biotechnology Center Total award amount: \$88,895 Total period covered: 7/01/2001-6/30/2003

Protein Hydrogen Bonding and NMR Redox Shifts of Cytochrome c Source: PRF Total award amount: \$60,000 Total period covered: 6/01/00-5/31/02

Replacement of a Failed Centrifuge Rotor Source: University Research Council Total award amount: \$4,000 Total period covered: 6/01/00-5/31/02

Bringing State-of-the-art NMR to UNC Source: University Research Council Total award amount: \$2,500 Total period covered: 1/01/00-12/31/01

Honors:

DuPont Young Faculty Award Morrow Young Faculty Award Folding & Binding Paper Alert selector for Current Opinions in Structural Biology, 1997-1998 Underwood Fund Award (BBSRC, U.K.) Invited Speaker, Proteins Gordon Conference, 2001 Invited Speaker, Biopolymers Gordon Conference, 2002, 2010 Invited Speaker, RASMB Gordon Conference, 2002 Invited Speaker FASEB Meeting: Protein Folding in the Cell, 2002 Invited Speaker, Toronto Chemical Biophysics Symposium, 2003, 2011 Invited Speaker 13th Conversation in Biomol. Stereodynamics, 2003 Invited Speaker, 18th Annual Gibbs Conference on Biothermodynamics, 2004 Invited Speaker, Eighth Johns Hopkins Folding Meeting, 2005 Invited Speaker, Colorado Protein Stability Conference, 2005 Invited Speaker, Cellular Osmoregulation: Sensors, Transducers & Regulators GRC, 2005 Invited Speaker Trends in Microcalorimetry 2005 Session Chair, Proteins GRC, 2005 Vice co-chair Proteins Gordon Research Conference, 2005 NIH Pioneer Award, 2006 Co-chair Proteins Gordon Research Conference, 2007 Invited Speaker, Southeast Magnetic Resonance Conference, 2007 Invited Speaker, Ions & Osmolytes Symposium, Salt Lake City ACS Meeting, 2008 Plenary Speaker, Beijing Conference & Exhibition on Instrumental Analysis, 2011 Program Committee Chair, 26th Annual Protein Society Symposium, 2012 Science Foundation of Ireland, E.T.S. Walton Visitor Award Invited Speaker, Molecular Crowding: Chemistry & Physics Meet Biology (Switzerland), 2012 Invited Speaker, 12th Chianti/INSTRUCT Workshop on BioNMR (Italy) 2012

Honors, continued

- Invited Speaker, EUROMAR (Dublin, Ireland), 2012
 - Glen H. Elder, Jr. Distinguished Term Professor of Research and Undergraduate Education, 2013-
 - Invited Speaker, Annual Protein Society Symposium, 2013
 - Invited Speaker, American Chemical Society National Meeting, New Orleans, April 7-11, 2013

Invited Speaker Graduate School Solvation Science Summer School, Bochum Germany, June 10-13, 2014

Invited Speaker Annual Meeting of the Biophysical Society of Japan. September 25-27, 2014

TC Wang Lecturer, Wuhan Institute of Physics and Mathematics, Chinese Academy of Sciences, March 18, 2015

Kenan Distinguished Professorship 2015-

2016 Chair, Biophysics In Vivo subgroup of the Biophysical Society

Invited Speaker, EMBO Conference on The Biochemistry and Chemistry of Biocatalysis: From Understanding to Design, Oulu, Finland, June 2016 Carl Brändén Award from the Protein Society, 2016

Invited Speaker, Intrinsically Disordered Proteins: Structure, Function & Interactions, Philadelphia, August 23

- Invited Speaker, Nobel Symposium on Protein Folding: From Mechanisms to Impact on Cells, Stockholm, Sweden, June 11-14, 2017
- University Mentor Award for Lifetime Achievement from UNC-CH. 2017

Invited Speaker, First International Symposium on Chemistry for Multimolecular Crowding Biosystems, Kobe, Japan, December 12-13, 2017

Invited Speaker, Physical and Quantitative Understanding of Cells at Molecular Level, Chemical Society of Japan, Kyoto, December 14-16, 2017

- Invited Speaker, Mini-workshop on Protein Biophysics: Interplay Between Experiments and Theories, Kyoto University, December 18, 2017
- Invited Speaker, EMBO Workshop, *In situ* methods in Cell Biology and Cellular Biophysics, Berlin, July 26-28, 2018
- Invited Speaker, Gibbs Conference on Biothermodynamics, Carbondale, IL, October 6-9, 2018
- Invited Speaker, 11th Biennial Carolina Biophysics Symposium, October 25, 2018
- Invited Speaker, Intrinsically Disordered Protein Subgroup Symposium, Biophysical Society Meeting, Baltimore, March 2, 2019
- Bradley University 2018 College of Liberal Arts and Sciences Distinguished Alumnus Award
- UNC-CH 2019 Excellence in Basic Science Mentoring Award

Invited Speaker, 20th Conversation in Biomolecular Structure and Dynamics, Albany, June 13 & 14, 2019 (2 talks)

Invited Speaker, Workshop on Macromolecular Crowding, Telluride, CO, July 16-20, 2019

- Invited Speaker, Workshop on Intrinsically-Disordered Proteins, Telluride, CO, July 16-20, 2019
- Invited Speaker, ACS Fall National Meeting, San Diego, August 25-29
- Invited Speaker, The Dr. and Mrs. Satti Paddi and Parvarti Reddy Public Lecture, Understanding Protein Behaviour in Living Cells, Memorial University, St. John's, Newfoundland, Canada, October 17
- Invited Speaker, Protein Folding Dynamics Gordon Research Conference, Galveston, TX, January 5-9, 2020

Plenary Lecture, EUROMAR, Portorož, Slovenia, remote, July 2021

Invited Speaker, Colorado Protein Stability Conference, Breckenridge, August 2022

Honors, continued

- Invited Speaker, German Biophysical Society (DGfB) Meeting Konstanz, Germany, September 2022
- Invited speaker, Specificity Determinants of Biomolecular Interactions, commemorating the late professor Aharon Katzir, Rehovot Israel, November 29, 2022
- UNC Johnston Teaching Excellence Award, 2023
- McElvian Lecture, Department of Chemistry, University of Wisconsin, Madison, February 21, 2023
- UNC Faculty Award for Excellence in Doctoral Mentoring, 2023
- Invited speaker, Prague Protein Spring meeting, May 4-6, 2023
- Invited speaker, Gesellschaft Deutscher Chemiker, Braunschweig, Germany, April 5, 2023
- Invited speaker, Telluride Workshop on Macromolecular Crowding, Telluride, CO, June 5-9, 2023
- Invited speaker, Biomolecules and Nanostructures meeting, Zakopane, Poland, June 14-18, 2023
- Fellow of the Biophysical Society, class of 2024
- Invited speaker, Protein Folding Dynamics Gordon Research Conference, Galveston, TX, January 7-12, 2024
- Plenary lecture, 20th European Magnetic Resonance Congress (Euromar), Bilbao, Spain June 30 – July 4

Research Seminars:

May 1, 2024-April 30, 2025

Plenary lecture, 20th European Magnetic Resonance Congress (Euromar), Bilbao, Spain June 30 – July 4

May 1, 2023-April 30, 2024

Prague Protein Spring meeting, May 4-6

Gesellschaft Deutscher Chemiker, Braunschweig, Germany, April 5 Telluride Workshop on Macromolecular Crowding, Telluride, CO, June 5-9 Biomolecules and Nanostructures meeting, Zakopane, Poland,

June 14-18

UC Berkeley, October 16

UCSF, October 18

Protein Folding Dynamics Gordon Research Conference, Galveston, TX,

January 8

Johns Hopkins, March 10

ACS Spring National Meeting, New Orleans, March 17-23

May 1, 2022-April 30, 2023

Colorado Protein Stability Conference, Breckenridge, August 9 ACS Fall National Meeting, August 21

German Biophysical Society (DGfB) Meeting Konstanz, Germany, September 20-23

Specificity Determinants of Biomolecular Interactions, commemorating the late professor Aharon Katzir, Rehovot Israel, November 29

McElvian Lecture, Department of Chemistry, University of Wisconsin, Madison, February 21

Department of Biochemistry and Biophysics, UNC-Chapel Hill, April 25

May 1, 2021-April 30, 2022

Plenary lecture, EUROMAR, Portorož, Slovenia (remote), July 4-8 Tulane, February 14 Institute of Pharmacology and Structural Biology, Toulouse (remote), March 15

May 1, 2020-April 30, 2021

ACS Fall National Meeting (virtual), August 17 BASF (virtual), September 23

May 1, 2019-April 30, 2020

20th Conversation in Biomolecular Structure and Dynamics, Albany, June 13 & 14 (2 talks)

Workshop on Macromolecular Crowding, Telluride, CO, July 16-20 Workshop on Intrinsically-Disordered Proteins, Telluride, CO, July 16-20 ACS Fall National Meeting, San Diego, August 25-29

The Dr. and Mrs. Satti Paddi and Parvarti Reddy Public Lecture, Understanding Protein Behaviour in Living Cells, Memorial University, St. John's, Newfoundland, Canada, October 17

Department of Chemistry, Memorial University, St. John's, Newfoundland, Canada, October 18

Protein Folding Dynamics Gordon Conference, Galveston, TX, January 5-9 University of Colorado, Denver, CO, March 6.

May 1, 2018-April 30, 2019

Suzhou Institute of Biomedical Engineering, Suzhou, China, May 10 Soochow University, Suzhou, China, May 11 Wuhan Institute of Physics and Mathematics, Chinese Academy of Sciences, Wuhan, China, May 17

- EMBO Workshop, In situ methods in Cell Biology and Cellular Biophysics, Berlin, July 26-28
- 2018 Gibbs Conference on Biothermodynamics, Carbondale, IL, October 6-9

Appalachian State University, Boone, NC, October 19

11th Biennial Carolina Biophysics Symposium, October 25

Bradley University, November 19

Weizmann Institute, Rehovot, Israel, January 29

- Fritz Haber Lecture, Hebrew University, Jerusalem, January 31
- 2019 Intrinsically Disordered Protein Subgroup Symposium, Biophysical Society Meeting, Baltimore, March 2, 2019
- Wuhan Institute of Physics and Mathematics, Chinese Academy of Sciences, Wuhan, China, March 27

May 1, 2017-April 30, 2018

China-Japan Joint Symposium on Functional Supramolecular Systems, Wuhan China, May 16

Wuhan Institute of Physics and Mathematics, Chinese Academy of Sciences, Wuhan, China, May 19

Nobel Symposium on Protein Folding: From Mechanisms to Impact on Cells, Stockholm, Sweden, June 11-14

New England Biolabs, December 7

First International Symposium on Chemistry for Multimolecular Crowding Biosystems, Kobe, Japan, December 12-13

Physical and Quantitative Understanding of Cells at Molecular Level, Chemical Society of Japan, Kyoto, December 14-16

Mini-workshop on Protein Biophysics: Interplay Between

Experiments and Theories, Kyoto University, December 18 Penn State University, February 22

UNC-CH, Department of Biochemistry and Biophysics, April 24

May 1, 2016-April 30, 2017

Appalachian State University, April 21 University of Wisconsin, March 3 Loyola University New Orleans, February 13 ACS Fall National Meeting, Philadelphia, August 21-25 Annual Protein Society Symposium, Baltimore, July 16-19 Ruhr-University Bochum, Germany, June 23 Leibniz-Institut für Molekulare Pharmakologie (FMP), Berlin, June 20 EMBO Conference on The Biochemistry and Chemistry of Biocatalysis: From Understanding to Design, Oulu, Finland, June 12-15

May 1, 2015-April 30, 2016

Higher Order Structure Conferences, Long Beach, CA, April 11-13 Hobart and William Smith Colleges, April 28

May 1, 2014-April 30, 2015

Workshop on Macromolecular Crowding, Telluride, CO, June 23-27 Washington Area NMR Group, December 4 California Separation Science Society, Higher Order Structure (CASSS-HOS) April 11-13

May 1, 2014-April 30, 2015

Graduate School Solvation Science Summer School, Bochum Germany, June 10-13 Novartis, Emeryville, July 22

Annual Meeting of the Biophysical Society of Japan, Sapporo, September 26 Tokyo Metropolitan University, September 30 University of North Carolina-Chapel Hill, October 8 University of Virginia, November 7 School of Life Sciences, U. of Science & Technology of China, Hefei, March 13 TC Wang Lecturer, Wuhan Institute of Physics and Mathematics, Chinese Academy of Sciences, March 18

May 1, 2013-April 30, 2014

Workshop on Macromolecular Crowding, Telluride, CO, June 25-29 Annual Protein Society Symposium, Boston, July 20-24 National Institute of Environmental Health Science, RTP, NC, September 5 University of Maryland, November 18 University of Puerto Rico, Rio Piedras Campus, November 22 Simon Fraser University, Vancouver, BC Canada, December 6 University of British Columbia, Vancouver, BC Canada, December 9 Florida State University, Tallahassee, January 21 Johns Hopkins, March 10 UNC Greensboro, March 28

May 1, 2012-April 30, 2013

University of Durham (UK), June 1 National University of Ireland, Galway, May 28 Molecular Crowding: Chemistry and Physics meet Biology (Switzerland), June 12 12th Chianti/INSTRUCT Workshop on BioNMR (Italy) June 18 EUROMAR (Dublin, Ireland), July 1 University of Tennessee, October 17 ACS Southeastern Regional Meeting (SERMACS). Raleigh, NC, November 16 Drexel University, February 18 University of Pennsylvania, Hershey, March 18 American Chemical Society National Meeting, New Orleans, April 7-11

May 1, 2011-April 30, 2012

Yale, March 19 King's College London, January 20 National Institute for Medical Research (UK), January 19 University of Oxford (UK), January 17 National Institutes of Health Pioneer Symposium, September 21 Beijing Conference & Exhibition on Instrumental Analysis, Oct 13 National Science Foundation, June 8 University of Minnesota Duluth, May 26

May 1, 2010-April 30, 2011

University of Toronto, June 3 Biopolymers Gordon Conference, June 6-11 IRB Barcelona, July 2 Swedish Royal Academy of Sciences, August 26 James Madison, September 3 Biological Diffusion & Brownian Dynamics Brainstorm 2 (Heidelberg, Germany) October 11 University of Indiana, October 25 National Science Foundation, November 10 University of Wisconsin, Madison, November 23 Toronto Chemical Biophysics Symposium, April 9

May 1, 2009 – April 30, 2010

University of Richmond, September 4 UCLA, March 11 Davidson, January 29

May 1, 2008 - April 30, 2009

Biophysical Society Workshop on Protein Folding, Stability, and Aggregation, Boston, March 3
Symposium on the Influence of Ions & Osmolytes on Aqueous Macromolecules, ACS Meeting, Salt Lake City, March 23
Workshop on Macromolecular Crowding, Telluride, CO, July 6-10
UNC Wilmington, September 28
Appalachian State University, November 21

May 1, 2007- April 30, 2008

UNC Chemistry, September 12 NIH Pioneer Symposium, September 19 University of Kansas, October 5 Southeastern Magnetic Resonance Conference, U. Alabama November 10 Honors Chemistry, UNC, November 19

May 1, 2006 – April 30, 2007

UNC, Chemistry, September 6
Duke, Biochemistry, October 2
University of Pennsylvania, Biophysics, October 18
Drexel University, Bioscience and Biotechnology, October 19
Virginia Tech, Chemistry, January 26
Biophysical Society, Intrinsically Disordered Proteins Subgroup, Baltimore, March 3
Seeing is Believing: The Future of Molecular and Biomolecular Imaging Meeting, Duke, March 11
UNC, Biochemistry & Biophysics, April 17

May 1, 2005 – April 30, 2006

Colorado Protein Stability Conference, Breckenridge, CO Trends in Microcalorimetry, Boston, MA

Cellular Osmoregulation: Sensors, Transducers & Regulators GRC, Newport, RI UNC Chemistry, Chapel Hill, NC UNC Biochemistry & Biophysics, Chapel Hill, NC Cold Spring Harbor Meeting on the Intracellular Molecular Environment, Cold Spring Harbor, NY University of Pittsburgh, Pittsburgh, PA University of Denver, Denver, CO University of Colorado, Health Sciences, Denver, CO U. Massachusetts, Amherst, MA NIH, Bethesda, MD

May 1, 2004 - April 30, 2005

Northern Illinois University, DeKalb, IL Rutgers University, New Jersey Johns Hopkins Folding Meeting, St. Michaels, MD Duke University, Durham University of Richmond, Chemistry

May 1, 2003 – April 30, 2004

Gibbs Conference, Carbondale, IL Microcalorimetry Conference, Atlanta Yale University, Molecular Biophys. & Biochemistry Emory University, Chemistry University of Kentucky, Biochemistry Wake Forest University, Physics UNC-Chapel Hill, Chemistry 13th Conversation in Biomolecular Stereodynamics, SUNY Albany

May 1, 2002 – April 30, 2003

Biopolymers Gordon Conference FASEB Protein Folding in the Cell Meeting Toronto Biophysics Symposium Rensselaer Polytechnic, Chemistry Penn. State, Chemistry Washington University, Biochemistry UNC-Chapel Hill, Biochemistry & Biophysics NC State University, Biochemistry UNC-Chapel Hill, Chemistry Drexel University, Biology University of Pennsylvania, Biophysics

May 1, 2001 – April 30, 2002

Proteins Gordon Conference Reversible Assoc. in Structural Molecular Biology Gordon Conference Boston ACI Proteomics Symposium University of Virginia, Biophysics Washington State University, Chemistry Sunesis, Inc., South San Francisco Stanford University, Biochemistry Georgia Tech., School of Chemistry and Biochemistry. Georgia State University, Chemistry

Patents:

Tardigrade disordered proteins as protein stabilizers US provisional patent application 62/375,238, Published March 25, 2022 Boothby T, Goldstein B, Pielak GJ, Piszkiewicz S, Brozena A

Device for particulate NMR samples in fluid US 8,773,130 B2 Pielak GJ, Barnes C, Sharaf N, Young G, Pinero F, Charlton L, Seagle C

Publications: [196, >12000 citations, H-index (Google Scholar) 64]

Brom JA, Samsri S, Petrikis RG, Parnham S, Pielak GJ. 2023. ¹H, ¹³C, ¹⁵N backbone resonance assignment of *Escherichia coli* adenylate kinase. Biomolecular NMR Assignments, in press.

Wang S, Eicher JE, Pielak GJ. 2023. Trifluoroethanol and the behavior of a tardigrade desiccation-tolerance protein. Protein Science, 32: e4716.

Eicher JE, Hutcheson, BO, Pielak G J. 2023. Properties of a tardigrade desiccation-tolerance protein aerogel. Biophysical Journal, 2500-2505.

Chu I-T, Pielak, GJ. 2023. Using NMR-detected hydrogen-deuterium exchange to quantify protein stability in cosolutes, under crowded conditions in vitro and in cells. Magnetic Resonance Letters 3, in press.

Chu I-T, Hutcheson BO, Malsch HR, Pielak GJ. 2023. Macromolecular crowding by polyethylene glycol reduces protein breathing. Journal of Physical Chemistry Letters, 14: 2599–2605.

Thole JF, Waudby C, Pielak GJ. 2023. Disordered proteins mitigate the temperature dependence of site-specific binding free energies. Journal of Biological Chemistry, 299: 102984.

Brom JA., Petrikis RG, Pielak GJ. 2023. How sugars protect dry protein structure. Biochemistry, 62: 1044-1052.

Stewart CJ, Olgenblum GI, Propst A, Harries D, Pielak GJ. 2023. Resolving the enthalpy of protein stabilization by macromolecular crowding. Protein Science, 32: e4573.

Eicher JE, Brom JA, Wang S, Sheiko SS, Atkin JM, Pielak GJ. 2022. Secondary structure and stability of a gel-forming tardigrade desiccation-tolerance protein. Protein Science, 31: e4495.

Zhang C, Pei Y, Zhang Z, Xu L, Liu X, Jiang L, Pielak GJ, Zhou X, Liu M, Li C. 2022. C-terminal truncation modulates α -synuclein's cytotoxicity and aggregation by promoting the interactions with membrane and chaperone. Communications Biology, 5: 798.

Brom J, Pielak GJ. 2022. Desiccation-tolerance- and globular- proteins adsorb similar amounts of water. *Protein Sci* 31: e4288.

Chu I-T, Stewart CJ, Speer SL, Pielak GJ. 2022. A difference between *in vitro* and in-cell protein dimer formation. *Biochemistry* 61:409-412.

Speer SL, Stewart C, Sapir L, Harries D, Pielak GJ. 2022. Macromolecular crowding is more than hard-core repulsions. Annual Review of Biophysics, 51: 267-300.

Crilly C, Brom JA, Warmuth O, Esterly HJ. 2022. Protection by desiccation-tolerance proteins probed at the residue level. Protein Science, 31: 396-406

Crilly C, Eicher JE, Warmuth O, Atkin JM, Pielak GJ. 2021. Water's variable role in protein

stability uncovered by liquid-observed vapor exchange NMR. Biochemistry, 60: 3041-3045.

Pielak GJ. 2021. Buffers, especially the good kind. Biochemistry, 60: 3436-3440.

Gruebele M, Pielak GJ. 2021. Dynamical spectroscopy and microscopy of proteins in cells. Current Opinion in Structural Biology, 70: 1-7.

Speer SL, Zheng W, Jiang X, Chu I-T, Guseman AJ, Liu M, Pielak GJ, Li C. 2021. The intracellular environment affects protein-protein interactions. *Proceedings of the National Academy of Sciences U S A* 118: e2019918118.

Thole J, Fadero T, Bonin J, Stadmiller S, Giudice J, Pielak G. 2021. *Danio rerio* oocytes for eukaryotic in-cell NMR. Biochemistry 60: 451-459.

Crilly C, Brom J, Kowalewski ME, Piszkiewicz S, Pielak, GJ. 2021. Dried protein structure revealed at the residue level by liquid-observed vapor exchange NMR. *Biochemistry* 60: 152-159.

Stadmiller SS, Pielak, GJ. 2020. Protein-complex stability in cells and in vitro under crowded conditions. *Current Opinion in Structural Biology*, 66: 183-192.

Stadmiller SS, Aguilar JS, Parnham S, Pielak GJ. 2020. Protein-peptide binding energetics under crowded conditions. *Journal of Physical Chemistry*, 42: 9297–9309.

Esterly, H. J.; Crilly, C. J.; Piszkiewicz, S.; Shovlin, D. J.; Pielak, G. J.; Christian, B. E. 2020. Toxicity and immunogenicity of a tardigrade cytosolic abundant heat soluble protein in mice. *Frontiers in Pharmacology*, 11, 565969.

Stadmiller SS, Aguilar, JS, Waudby C, Pielak GJ. 2020. Rapid quantification of protein-ligand binding via ¹⁹F NMR lineshape analysis. *Biophysical Journal*, 118: 2333-2335.

Chu I-T, Speer SL, Pielak GJ. 2020. Rheostatic control of protein expression using Tuner cells. *Biochemistry*, 59: 733-735.

Pielak G, Piszkiewicz S 2019. Protecting enzymes from stress-induced inactivation *Biochemistry* 58: 3825-3833.

Ye Y, Wu Q, Zheng W, Jiang B, Pielak G, Liu M, Li C. 2019. Positively-charged tags impede protein mobility in cells as quantified by ¹⁹F NMR. *Journal of Physical Chemistry*, 123: 4527-4533.

Speer SL, Guseman AJ, Patteson JB, Ehrmann BM, & Pielak GJ. 2019. Controlling and quantifying protein concentration in *Escherichia coli*. *Protein Science* 28: 1307-1311.

Jiang X, Zhang Z, Cheng K, Wu Q, Jiang L, Pielak GJ, Liu M, & Li C. 2019. Membrane-mediated disorder-to-order transition of SNAP25 flexible linker facilitates its interaction with syntaxin-1 and snare-complex assembly. *The FASEB Journal*, 33: 7985-7994.

Piszkiewicz S, Gunn KH, Warmuth O, Propst A, Mehta A, Nguyen KH, Kuhlman E, Guseman AJ, Stadmiller SS, Boothby TC, Neher SB, & Pielak GJ. 2019. Protecting activity of desiccated enzymes. *Protein Science* 28, 941-951.

Carter C, Wolfenden R, Caplow M, Lentz B, Pielak G, Watenpaugh K, Hu H, & Puett D. 2019. Jan Hermans (1933-2018): Red-blooded biophysicists study hemoglobin. *Proteins* 87:171-173.

Stadmiller SS, Pielak GJ. 2018. The expanding zoo of in-cell protein NMR. *Biophysical Journal*. 115: 1628-1629

Cheng K, Wu Q, Zhang Z, Pielak GJ, Liu M, Li C. 2018. Crowding and confinement can oppositely affect protein stability. ChemPhysChem 19: 1-7.

Guseman AJ, Perez Goncalves GM, Speer SL, Young GB, Pielak GJ. 2018. Protein shape modulates crowding effects. *Proceedings of the National Academy of Sciences of the United States of America*, 115: 10965-10970.

Stadmiller SS Pielak GJ. 2018. Enthalpic stabilization of an SH3 domain by D₂O. *Protein Science*, 27: 1710-1716.

Rydeen AE, Brustad EM, Pielak GJ. 2018. Osmolytes and Protein–Protein interactions. *Journal of the American Chemical Society*. 140: 7441-7444.

Guseman SJ, Speer SL, Perez Goncalves GM, Pielak GJ. 2018. Surface charge modulates proteinprotein interactions in physiologically relevant environments. *Biochemistry*, 57: 1681-1684.

Ye Y, Wu Q, Zheng W, Jiang B, Pielak GJ, Liu M, Li C. 2017. Quantification of size effect on protein rotational mobility in cells by ¹⁹F NMR spectroscopy. *Analytical and Bioanalytical Chemistry*, 410: 869-874.

Acosta LC, Perez Goncalves GM, Pielak GJ, Gorensek-Benitez AH. 2017. Large cosolutes, small cosolutes and dihydrofolate reductase activity. *Protein Science*, 26: 2417–2425.

Boothby TC, Pielak GJ. 2017. Intrinsically disordered proteins and desiccation tolerance: elucidating functional and mechanistic underpinnings of anhydrobiosis. *BioEssays*, 39 (11).

Gorensek-Benitez AH, Smith AE, Stadmiller SS, Perez Goncalves GM, Pielak GJ. 2017. Cosolutes, crowding, and protein folding kinetics. *Journal of Physical Chemistry B*, 121: 6527-6537.

Cohen RD, Pielak GJ 2017. Protein quinary interactions with an unfolded state ensemble. *Protein Science*, 26: 1698-1703.

Boothby TC, Tapia H, Brozena AH, Piszkiewicz S, Smith AE, Giovannini I, Rebecchi L, Pielak GJ, Koshland D, & Goldstein B. 2017. Tardigrades use intrinsically disordered proteins to survive desiccation. *Molecular Cell*, 65: 975–984. *Featured by National Public Radio, The BBC, The New York Times, The Christian Science Monitor, The Telegraph, The New Scientist, Chemical and Engineering News, Trends in Biochemical Sciences and the Faculty of 1000.*

Guseman, GJ, Pielak GJ 2017. Cosolute and crowding effects on a side-by-side protein dimer. *Biochemistry*, 56: 971–976.

Li C, Zhao J, Zhang X, Zhou X, Pielak GJ, Liu M, Cheng K, Ge Y, Wu Q, Ye Y, Xu G, Zhang Z, Zheng W 2017. Magnetic resonance spectroscopy as a tool for assessing macromolecular structure and function in living cells. *Annual Review of Analytical Chemistry* 10: 157-182.

Cohen RD, Pielak GJ 2017. A cell is more than the sum of its (dilute) parts: a brief history of quinary structure. *Protein Science* 26: 403-413.

Bia J, Pielak GJ, Li C 2017. Macromolecular and small molecular crowding have similar effects on α-synuclein structure. *ChemPhysChem*, 18, 55-58.

Cohen RD, Pielak GJ 2016. Electrostatic contributions to protein quinary structure. *Journal of the American Chemical Society* 138: 13139–13142.

Senske M, Smith A, Pielak GJ 2016. Protein stability in reverse micelles. *Angewandte Chemie* 55, 3586-3589.

Smith AE, Zhou LZ, Gorensek AH, Senske M, Pielak GJ 2016. In-cell thermodynamics and a new role for protein surfaces. *Proceedings of the National Academy of Sciences of the United States of America*, 113, 1725-1730. Featured by Faculty of 1000.

Zhaia Z, Wua Q, Zheng W, Liu M, Pielak GJ, Li C 2016. Roles of structural plasticity in chaperone HdeA activity are revealed by ¹⁹F NMR. *Chemical Science*, *7*, 2222–2228.

Tyrrell J, Weeks, KM, Pielak GJ 2015. Challenge of mimicking the influences of the cellular environment on RNA structure by PEG-induced macromolecular crowding. *Biochemistry*, 54: 6447–6453. **Featured by Faculty of 1000.**

Cohen RD, Guseman AJ, Pielak GJ 2015. Intracellular pH modulates quinary structure. *Protein Science*, 24: 1748–1755.

Spitzer J, Pielak GJ, Poolman B. 2015. Emergence of life: Physical chemistry changes the paradigm. *Biology Direct*, 10: 33.

Smith AE, Zhou Z, Pielak GJ. 2015. Hydrogen exchange of disordered proteins in *Escherichia coli*. *Protein Science*, 24: 706-713. **Featured by Faculty of 1000.**

Monteith WB, Cohen RD, Smith AE, Guzman-Cisneros E, Pielak GJ. 2015. Quinary structure modulates protein stability in cells. *Proceedings of the National Academy of Sciences of the United States of America*, 112: 1739-1742. **Featured in C&EN Concentrates.**

Smith AE, Zhang Z, Pielak GJ, Li C 2015. NMR studies of protein folding and binding in cells and cell-like environments. *Current Opinion in Structural Biology*. 30: 7-16.

Sarkar M, Pielak GJ 2014. An osmolyte mitigates the destabilizing effect of protein crowding. *Protein Science* 23: 1161-1164. **Cover article.**

Monteith WB, Pielak GJ 2014. Residue level quantification of protein stability in living cells. *Proceedings of the National Academy of Sciences of the United States of America* 111: 11335-11340. Featured by Faculty of 1000.

Theillet F-X, Binolfi A, Frembgen-Kesner T, Hingorani K, Sarkar M, Kyne C, Li C, Crowley P, Gierasch L, Pielak G, Elcock A, Gershenson A, Selenko P. 2014. Physicochemical properties of cells and their effects on intrinsically disordered proteins (IDPs). *Chemical Reviews* 13: 6661-6714.

Xu G, Ye Y, Liu X, Cao S, Wu Q, Cheng K, Liu M, Pielak GJ, Li C. 2014. Strategies for Protein NMR in *Escherichia coli*. *Biochemistry* 53: 1971-1981.

Sarkar M, Lu J, Pielak GJ. 2014. Protein-crowder charge and protein stability. *Biochemistry* 53: 1601-1606.

Tyrrell J, McGinnis JL, Weeks KM, Pielak GJ. 2013. The cellular environment stabilizes adenine riboswitch RNA structure. *Biochemistry*, 52: 8777–8785. Featured by Faculty of 1000.

Sarkar M, Smith AE, Pielak GJ. 2013. Impact of reconstituted cytosol on protein stability. *Proceedings of the National Academy of Sciences of the United States of America* 110: 19342-19347. **Featured by Faculty of 1000.**

Smith AE, Sarkar M, Young GB, Pielak GJ. 2013. Amide proton exchange of a dynamic loop in cell extracts. *Protein Science* 22: 1313-1319.

Ye Y, Liu X, Zhang Z, Wu Q, Jiang B, Jiang L, Zhang X, Liu M, Pielak GJ, C Li C. 2013. ¹⁹F

NMR as a probe of cytoplasmic viscosity and weak protein interactions in living cells. *Chemistry--A European Journal* 19: 12705-12710.

Sarkar M, Li C, Pielak GJ. 2013. Soft interactions and crowding. *Biophysical Reviews* 5: 187-194.

Benton LA, Smith AE, Young GB, Pielak GJ. 2012. Unexpected effects of macromolecular crowding on protein stability. *Biochemistry* 51: 9773-9775.

Wang Y, Sarkar M, Smith AE, Krois AS, Pielak, GJ. 2012. Macromolecular crowding and protein stability. *Journal of the American Chemical Society* 134: 16614–16618.

Wang Y, Benton L, Singh V, Pielak GJ. 2012. Disordered protein diffusion under crowded conditions. *Journal of Physical Chemistry Letters* 3: 2703–2706.

Wang Y, Li C, Pielak GJ. 2012. In-cell protein magnetic resonance spectroscopy. *Chinese Journal of Magnetic Resonance*, 29: 475-488.

Pielak GJ, Tian F. 2012. Membrane proteins, magic-angle spinning, and in-cell NMR. *Proceedings of the National Academy of Sciences of the United States of America* 109: 4715-4716.

Zigoneanu IG, Pielak GJ 2012. Interaction of α -synuclein and a cell penetrating fusion peptide with higher eukaryotic cell membranes assessed by ¹⁹F NMR. *Molecular Pharmaceutics* 9: 1024-1029.

Zigoneanu IG, Yang YJ, Krois AS, Haque Md E, Pielak GJ. 2011. Interaction of α -synuclein and its A30P variant with vesicles of composition similar to mitochondrial membranes. *Biochimica et Biophysica Acta* 1818: 512-519.

Fu R, Wang X, Li C, Santiago-Miranda A, Pielak GJ, Tian F. 2011. *In situ* Structural characterization of a recombinant protein in native *Escherichia coli* membranes with solid-state MAS NMR. *Journal of the American Chemical Society* 133: 12370-12373.

Schlesinger AP, Wang Y, Tadeo X, Millet O, Pielak GJ. 2011 Macromolecular crowding fails to fold a globular protein in cells. *Journal of the American Chemical Society* 133: 8082-8085.

Miklos AC, Sarkar M, Wang Y, Pielak GJ. 2011. Protein crowding tunes protein stability. *Journal of the American Chemical Society* 133: 7116–7120. Featured by Faculty of 1000.

Miklos AC, Li C, Sorrell CD, Lyon LA, Pielak GJ. 2011. An upper limit for macromolecular crowding effects. *BMC Biophysics* 4:13.

Barnes CO, Monteith WB, Pielak GJ. 2011. Internal and global protein motion assessed with a fusion construct and in-cell NMR. *ChemBioChem* 12: 390-391.

Barnes CO, Pielak GJ. 2011. In-cell NMR and protein leakage. *Proteins: Structure, Function, and Bioinformatics* 79: 347-351.

Miklos AC, Pielak GJ 2010. Crowding and function reunite. *Proceedings of the National Academy of Sciences of the United States of America* 107: 17457-17458.

Li C, Wang G-F, Pielak GJ 2010. Probing the micelle-bound aggregation-prone state of α -synuclein with ¹⁹F NMR. *ChemBioChem* 11: 1993-1996.

Miklos AC, Li C, Sharaf NG, Pielak GJ 2010. Volume exclusion and soft interaction effects on

protein stability under crowded conditions. Biochemistry 49: 6894-6991.

Wang G-F, Li C, Pielak GJ. 2010. ¹⁹F NMR studies of α-synuclein-membrane interactions. *Protein Science* 19: 1686-1691.

Wang Y, Li C, Pielak GJ. 2010. Effects of proteins on protein diffusion. *Journal of the American Chemical Society* 132: 9392-9397.

Li C, Wang G-F, Wang Y, Creager-Allen R, Lutz EA, Scronce H, Slade K M. Ruf RA, Mehl RA Pielak GJ 2010. Protein ¹⁹F NMR in *Escherichia coli*. *Journal of the American Chemical Society* 132: 321-327. Featured in *C&EN* Concentrates.

Sharaf NG, Barnes CO, Charlton LM, Young GB, Pielak GJ. 2010. A bioreactor for in cell protein NMR. *Journal of Magnetic Resonance*: 202: 140-146. **Cover article.**

Miklos AC, Li C, Pielak GJ. 2009. Using NMR-detected backbone amide ¹H exchange to assess macromolecular crowding effects on globular-protein stability. *Methods in Enzymology* 466: 1-18.

Li C, Wang Y, Pielak GJ. 2009. Translational and rotational diffusion of a small globular protein under crowded conditions. *Journal of Physical Chemistry B* 113: 13390–13392.

Li C, Lutz EA, Slade KM, Ruf RA, Wang G, Pielak GJ. 2009. ¹⁹F-NMR studies of asynuclein conformation and fibrillation. *Biochemistry* 48: 8578–8584.

Slade KM, Baker R, Chua M, Thompson NL, Pielak GJ. 2009. Effects of recombinant protein expression on green fluorescent protein diffusion in *Escherichia coli*. *Biochemistry* 48: 5083–5089.

Slade KM, Steele BL, Pielak GJ, Thompson NL. 2009. Quantifying GFP diffusion in *Escherichia coli* by using continuous photobleaching with evanescent illumination. *Journal of Physical Chemistry* 113: 4837-4845.

Li C, Pielak GJ 2009. Using NMR to distinguish viscosity effects from nonspecific protein binding under crowded conditions. *Journal of the American Chemical Society* 131: 1368-1369.

Pielak GJ, Li C, Miklos AC, Schlesinger AP, Slade K M, Wang G., Zigoneanu IG. 2009. Protein NMR under physiological conditions. *Biochemistry* 48: 226–234.

Ruf RA, Lutz EA, Zigoneanu IG, Pielak G J. 2008. α -Synuclein conformation affects its tyrosine-dependent oxidative aggregation. *Biochemistry* 47: 13604–13609.

Li C, Charlton LM, Lakkavaram A, Seagle C, Wang G, Young GB, Macdonald JM, Pielak GJ. 2008. Differential dynamical effects of macromolecular crowding on an intrinsically disordered protein and a globular protein: implications for in-cell NMR. *Journal of the American Chemical Society* 130: 6310-6311.

Charlton LM, Barnes CO, Li C, Orans J, Young GB, Pielak GJ. 2008. Residue-level interrogation of macromolecular crowding effects on protein stability. *Journal of the American Chemical Society* 130: 6826-6830.

Pielak GJ, Patel CN, Winzor DJ. 2007. Reconsideration of sedimentation equilibrium distributions reflecting the effects of small inert cosolutes on the dimerization of alphachymotrypsin. *Biophysical Chemistry* 130: 89-92. Charlton LM, Pielak GJ 2006. Peeking into living eukaryotic cells with high-resolution NMR, *Proceedings of the National Academy of Sciences of the United States of America* 103: 11817-11818.

Bryant JE, Lecomte JTJ, Lee AL, Young GB, Pielak GJ 2006. Cytosol has a small effect on protein backbone dynamics. Biochemistry 45: 10085-10091. Retracted: *ibid*. 46: 8206.

McCall S J, Nassar R, Malouf NN, Saunders AJ, Oakeley, AE, Henderson PM, Solaro RJ, Pielak GJ, Alexander KA, and Anderson PAW 2006. Development and cardiac contractility: cardiac troponin T isoforms and cytosolic calcium. *Pediatric Research* 60: 276-281.

Pielak GJ. 2006. Woes of proline: a cautionary kinetic tale. Protein Science 15: 393-394.

McNulty BC, Young GB, Pielak GJ. 2006. Macromolecular crowding in the *Escherichia coli* periplasm maintains a-synuclein disorder. *Journal of Molecular Biology* 355: 893-897.

McNulty BC, Tripathy A, Young GB, Orans J, Pielak GJ. 2006. Temperature-induced reversible conformational change in the first 100 residues of a-synuclein. *Protein Science* 15: 602-608.

Barrett DG, Minder CM, Mian MU, Whittington SJ, Cooper J, Fuchs KM, Tripathy A, Waters ML, Creamer TP, Pielak GJ. 2006. Pressure perturbation calorimetry of helical peptides. *Proteins: Structure Function and Bioinformatics*. 63: 322-326.

Daughdrill GW, Pielak GJ, Uversky VN, Cortese MS, Dunker AK. 2005. In *Protein Folding Handbook*; Buchner J, Kiefhaber T, Eds.; Wiley-VCH: Weinheim, p 275-357.

Bryant JE, Lecomte JTJ, Lee AL, Young GB, Pielak GJ. 2005. Protein dynamics in living cells. *Biochemistry* 44: 9275-9279. Retracted: *ibid*. 46: 8206.

Pielak GJ 2005. A model of cellular organization. *Proceedings of the National Academy of Sciences of the United States of America* 102: 5901-5902.

Yi F, Sims D, Pielak GJ, Edgell MH. 2005. The impact of robotics and instrument automation on protein stability measurements. *Journal of the Association for Laboratory Automation* 10: 98-101.

Olteanu A, Pielak GJ. 2004. Peroxidative aggregation of α -synuclein requires tyrosines. *Protein Science* 13: 2852-2856.

Batchelor JD, Olteanu A, Tripathy A, Pielak GJ 2004. Impact of protein denaturants and stabilizers on water structure. *Journal of the American Chemical Society* 126: 1958-1961. **Featured by Faculty of 1000.**

Olteanu A, Patel CN, Dedmon MM, Kennedy S, Linhoff MW, Minder CM, Potts PR, Deshmukh M, Pielak GJ 2003. Stability and apoptotic activity of recombinant human cytochrome *c. Biochemical and Biophysical Research Communications* 312/3: 733-740.

Yi F, Sims DA, Pielak GJ, Edgell MH. 2003. Testing hypotheses about determinants of protein structure with high-precision high-throughput stability measurements and statistical modeling. *Biochemistry* 42: 7594-7603.

Edgell MH, Sims DA, Pielak GJ, Yi F. 2003. High-precision high-throughput stability determinations facilitated by robotics and a semi-automated titrating fluorometer. *Biochemistry* 42: 7587-7593.

Engstrom G, Rajagukguk R, Saunders AJ, Patel CN, Rajagukguk S, Merbitz-Zahradnik T, Xiao K, Pielak GJ, Trumpower B, Yu C-A, Yu L, Durham B, Millett F. 2003. Design of a ruthenium-labeled cytochrome *c* derivative to study electron transfer with the cytochrome *bc1* complex. *Biochemistry* 42: 2816-2824.

Dedmon MM, Patel CN, Young GB, Pielak GJ. 2002. FlgM gains structure in living cells. *Proceedings of the National Academy of Sciences of the United States of America* 99: 12861-12864. **Featured by Faculty of 1000.**

Beasley JR, Doyle DF, Chen L, Cohen DS, Fine BR, Pielak GJ. 2002. Searching for quantitative entropy-enthalpy compensation among protein variants. *Proteins: Structure, Function, and Genetics* 49: 398-402.

Patel CN, Noble S., Weatherly GT, Tripathy A, Winzor DJ, Pielak GJ. 2002. Effects of molecular crowding by saccharides on alpha-chymotrypsin dimerization. *Protein Science* 11: 997-1003.

Morar AS, Pielak GJ. 2002. Crowding by trisaccharides and the 2:1 cytochrome *c*/cytochrome *c* peroxidase complex. *Biochemistry* 41: 547-551.

Morar AS, Olteanu A, Young GB, Pielak GJ. 2001. Solvent-induced collapse of alphasynuclein and acid denatured cytochrome *c. Protein Science* 10: 195-199.

Patel C, Lind M, Pielak GJ. 2001. Characterization of horse cytochrome *c* expressed in *Escherichia coli*. *Protein Expression and Purification:* 22: 220-224.

Davis-Searles PR, Saunders AJ, Erie DA, Winzor DJ, Pielak GJ. 2001. Interpreting the effects of small uncharged solutes on protein-folding equilibria. *Annual Review of Biophysics and Bioengineering* 30: 271-306.

Morar AS, Wang X, Pielak GJ. 2001. Effects of crowding by mono-, di-, and tetrasaccharides on cytochrome *c* / cytochrome *c* peroxidase binding: comparing theory to experiment. *Biochemistry* 40: 281-285.

Pielak GJ, Wang X. 2001. Interactions between yeast iso-1-cytochrome *c* and its peroxidase. *Biochemistry* 40: 422-428.

Weatherly GT, Pielak GJ. 2001. Second virial coefficients as a measure of proteinosmolyte interactions. *Protein Science* 10: 12-16.

Saunders AJ, Davis-Searles PR, Allen DL, Pielak GJ, Erie DA. 2000. Osmolyte-induced changes in protein conformational equilibria. *Biopolymers* 53: 293-307.

Boyd J, Dobson CM, Morar AS, Williams RJP, Pielak GJ. 1999. ¹H and ¹⁵N hyperfine shifts of cytochrome *c. Journal of the American Chemical Society* 121: 9247-9248.

Chen L, Pielak GJ, Thompson NL. 1999. The cytoplasmic region of $Fc(\gamma)$ RIIb1, but not $Fc(\gamma)$ RIIb2, binds phospholipids membranes. *Biochemistry* 38: 2102-2109.

Hostetter DR, Weatherly GT, Beasley JR, Bortone K, Cohen DS, Finger SA, Hardwidge P, Kakouras D, Saunders AJ, Trojak SK, Waldner JL, Pielak GJ. 1999. Partially formed

native tertiary interactions in the A-state of cytochrome *c. Journal of Molecular Biology* 289: 639-644.

Lahr SJ, Broadwater A, Carter CW, Jr., Collier ML, Hensley L, Waldner JL, Pielak GJ, Edgell MH. 1999. Patterned library analysis: a method for the quantitative assessment of hypotheses concerning the determinants of protein structure. *Proceedings of the National Academy of Sciences of the United States of America* 96: 14860-14865.

Mei H, Wang K, Peffer N, Weatherly GT, Cohen DS, Pielak GJ, Durham B, Millett F. 1999. Role of configurational gating in intramolecular electron transfer from cytochrome *c* to the radical cation in cytochrome *c* peroxidase. *Biochemistry* 39: 6846-6854.

Morar AS, Kakouras D, Young GB, Boyd J, Pielak GJ. 1999. Expression of ¹⁵N-labeled eukaryotic cytochrome *c* in *Escherichia coli*. *Journal of Biological Inorganic Chemistry* 4: 220-222.

Waldner JL, Lahr SJ, Edgell MH, Pielak GJ. 1999. Nonideality and protein thermal denaturation. *Biopolymers* 49: 471-479.

Wang X, Pielak GJ. 1999. Equilibrium thermodynamics of a physiologically-relevant heme-protein complex. *Biochemistry* 38: 16876-16881.

Allen DL, Pielak GJ. 1998. Baseline length and automated fitting of denaturation data. *Protein Science* 7: 1262-1263.

Davis-Searles PR, Morar AS, Saunders AJ, Erie DA, Pielak GJ. 1998. Sugar-induced molten-globule model. *Biochemistry* 37: 17048-17053.

Fairris JL, Wang K, Geren L, Pielak GJ, Durham B, Millett F. 1998. Intramolecular electron transfer in yeast cytochrome *c* covalently bonded to ruthenium(II) polypyridine complexes at cys39. In: *Photochemistry and Radiation Chemistry: American Chemical Society Advances in Chemistry Series 254*. American Chemical Society: Washington DC pp. 99-110.

Marmorino JL, Lehti M, Pielak GJ. 1998. Native tertiary structure in an A-state. *Journal of Molecular Biology* 275: 379-388.

Waldner JL, Lahr SJ, Edgell MH, Pielak GJ. 1998. Effects of a polyhistidine terminal extension on eglin c stability. *Analytical Biochemistry* 263: 116-118.

Wrobel JA, Chao S-F, Conrad MJ, Merker JD, Swanstrom R, Pielak GJ, Hutchison CA, III. 1998. A genetic approach for identifying critical residues in the fingers and palm subdomains of HIV-1 reverse transcriptase. *Proceedings of the National Academy of Sciences of the United States of America* 95: 638-645.

Chen L, Thompson NL, Pielak GJ. 1997. Design, synthesis, and characterization of the genes for mouse Fc γ RIIb1 and Fc γ RIIb2 cytoplasmic regions. *Protein Science* 6: 1038-1046.

Pielak GJ. 1997. Review of "Circular dichroism and the conformational analysis of biomolecules." Fasman G.D., Ed., Plenum NY (1997). *American Scientist* 85: 391-392.

Beasley JR, Pielak GJ. 1996. Requirements for perpendicular helix pairing. *Proteins: Structure Function and Genetics* 26: 95-107.

Betz, SF, Marmorino JL, Saunders AJ, Doyle DF, Young GB, Pielak GJ. 1996. Unusual effects of an engineered disulfide on global and local protein stability. *Biochemistry* 35: 7422-7428.

Doyle DF, Waldner JL, Parikh S, Alcazar-Roman L, Pielak GJ. 1996. Changing the transition state for protein (un)folding. *Biochemistry* 35: 7403-7411.

Mei H, Wang K, McKee S, Wang X, Waldner JL, Pielak GJ, Durham B, Millett F. 1996. Control of formation and dissociation of the high-affinity complex between cytochrome *c* and cytochrome *c* peroxidase by ionic strength and the low-affinity binding site. *Biochemistry* 35: 15800-15806.

Miller MA, Geren L, Han GW, Saunders A, Beasley J, Pielak GJ, Durham B, Millett F, Kraut J. 1996. Identifying the physiological electron transfer site of cytochrome *c* peroxidase by structure-based engineering. *Biochemistry* 35: 667-673.

Pappa HS, Tajbaksh S, Saunders AJ, Pielak GJ, Poulos TL. 1996. Probing the cytochrome *c* peroxidase-cytochrome *c* electron transfer reaction using site specific crosslinking. *Biochemistry* 35: 4837-4845.

Pielak GJ, Auld DS, Betz SF, Hilgen-Willis SE, Garcia LL. 1996. Nuclear magnetic resonance studies of class I cytochromes *c*. In: Scott RA, Mauk AG, eds. *Cytochromes c: A Multidisciplinary Approach*. Sausalito: University Science Books. pp. 203-284.

Wang K, Mei H, Geren L, Miller MA, Saunders A, Wang X, Waldner JL, Pielak GJ, Durham B, Millett F. 1996. Design of a ruthenium-cytochrome *c* derivative to measure electron transfer to the radical cation and oxyferryl heme in cytochrome *c* peroxidase. *Biochemistry* 35: 15107-15119.

Cohen DS, Pielak GJ. 1995. Entropic stabilization of cytochrome *c* upon reduction. *Journal of the American Chemical Society* 117: 1675-1677.

Geren LM, Beasley JR, Fine BR, Saunders AJ, Hibdon S, Pielak GJ, Durham B, Millett F. 1995. Design of a ruthenium-cytochrome *c* derivative to measure electron transfer to the initial acceptor in cytochrome *c* oxidase. *Journal of Biological Chemistry* 270: 2466-2472.

Marmorino JL, Pielak GJ. 1995. A native tertiary interaction stabilizes the A state of cytochrome *c. Biochemistry* 34: 3140-3143.

Pielak GJ, Auld DS, Beasley JR, Betz, SF, Cohen DS, Doyle DF, Finger SA, Fredericks ZL, Hilgen-Willis S, Saunders AJ, Trojak SK. 1995. Protein thermal denaturation, side-chain models, and evolution: amino acid substitutions at a conserved helix-helix interface. *Biochemistry* 34: 3268-3276.

Cohen DS, Pielak GJ. 1994. The stability of yeast iso-1-cytochrome *c* as a function of pH and temperature. *Protein Science* 3: 1253-1260.

Auld DS, Young GB, Saunders AJ, Doyle DF, Pielak GJ. 1993. Probing weakly-polar interactions in cytochrome *c. Protein Science* 2: 2187-2197.

Fredericks ZL, Pielak GJ. 1993. Exploring the interface between the N- and C-terminal helices of cytochrome *c* by random mutagenesis within the C-terminal helix. *Biochemistry* 32: 929-936.

Hilgen-Willis S, Bowden EF, Pielak GJ. 1993. Dramatic stabilization of ferricytochrome *c* upon reduction. Journal of Inorganic *Biochemistry* 51: 649-653.

Marmorino JL, Auld DS, Betz, SF, Doyle DF, Young GB, Pielak GJ. 1993. Amide proton exchange rates of oxidized and reduced *Saccharomyces cerevisiae* iso-1-cytochrome *c*. Protein Science 2: 1966-1974.

Saunders AJ, Young GB, Pielak GJ. 1993. Polarity of disulfide bonds. *Protein Science* 2: 1183-1184.

Greene RM, Betz, SF, Hilgen-Willis S, Auld DS, Fencl JB, Pielak GJ. 1993. Changes in global stability and local structure of cytochrome *c* upon substituting phenylalanine-82 with tyrosine. *Journal of Inorganic Biochemistry* 51: 663-676.

Willie A, McLean M, Liu R-Q, Hilgen-Willis S, Saunders AJ, Pielak GJ, Sligar SG, Durham B, Millett F. 1993. Intracomplex electron transfer between ruthenium-65-cytochrome b_5 and position-82 variants of yeast iso-1-cytochrome *c. Biochemistry* 32: 7519-7525.

Betz, SF, Pielak GJ. 1992. Introduction of a disulfide bind into cytochrome *c* stabilizes a compact denatured state. *Biochemistry* 31: 12337-12344.

Gao Y, McLendon G, Pielak GJ, Williams RJP. 1992. Electron-proton coupling in cytochrome *c* studied using protein variants. *European Journal of Biochemistry* 204: 337-352.

Auld DS, Pielak GJ. 1991. Constraints on amino acid substitutions in the N-terminal helix of cytochrome *c* explored by random mutagenesis. *Biochemistry* 30: 8684-8690.

Concar DW, Whitford D, Pielak GJ, Williams RJP. 1991. The role of phenylalanine-82 in electron-exchange reactions of eukaryotic cytochromes *c. Journal of the American Chemical Society* 113: 2401-2406.

Gao Y, Boyd J, Pielak GJ, Williams RJP. 1991. Proton nuclear magnetic resonance as a probe of differences in structure between the C102T and F82S;C102T variants of iso-1-cytochrome *c* from the yeast *Saccharomyces cerevisiae*. *Biochemistry* 30: 7033-7040.

Gao Y, Boyd J, Pielak GJ, Williams RJP. 1991. Comparison of reduced and oxidized yeast iso-1-cytochrome *c* using proton paramagnetic shifts. *Biochemistry* 30: 1928-1934.

Hilgen SE, Pielak GJ. 1991. The function of the *Saccharomyces cerevisiae* iso-1-cytochrome *c* gene is independent of the codon at invariant residue phe82 when the gene is present on a low copy number vector. *Protein Engineering* 4: 575-578.

Thurgood AGP, Pielak GJ, Cutler RL, Davies AM, Greenwood C, Mauk AG, Smith M, Williamson DJ, Moore GR. 1991. Change in charge of an unvaried heme contact residue does not cause a major change of conformation in cytochrome *c. Federation of European Biochemical Societies Letters* 284: 173-177.

Wang X, Pielak GJ. 1991. Temperature-sensitive variants of *Saccharomyces cerevisiae* iso-1- cytochrome *c* produced by random mutagenesis of codons 43 to 54. *Journal of Molecular Biology* 221: 97-105.

Hildebrandt P, Pielak GJ, Williams RJP. 1991. Structural studies of yeast iso-1cytochrome c mutants by resonance Raman spectroscopy. *European Journal of Biochemistry* 201: 211-216.

Gao Y, Boyd J, Williams RJ, Pielak GJ. 1990. Assignment of proton resonances, identification of secondary structural elements, and analysis of backbone chemical shifts

for the C102T variant of yeast iso-1-cytochrome *c* and horse cytochrome *c*. *Biochemistry* 29: 6994-7003.

Liang N, Mauk AG, Pielak GJ, Johnson JA, Smith M, Hoffman BM. 1988. Regulation of interprotein electron transfer by residue 82 of yeast cytochrome *c. Science* 240: 311-313.

Pielak GJ, Atkinson RA, Boyd J, Williams RJP. 1988. Two-dimensional NMR as a probe of structural similarity applied to mutants of cytochrome *c. European Journal of Biochemistry* 177: 179-185.

Pielak GJ, Boyd J, Moore GR, Williams RJP. 1988. Proton NMR studies show that the THR 102 mutant of yeast iso-1-cytochrome *c* is a typical member of the cytochrome *c* family. *European Journal of Biochemistry* 177: 167-177.

Cutler RL, Pielak GJ, Mauk AG, Smith M. 1987. Replacement of cysteine-107 of *Saccharomyces cerevisiae* iso-1- cytochrome *c* with threonine: improved stability of the mutant protein. *Protein Engineering* 1: 95-99.

Liang N, Pielak GJ, Mauk AG, Smith M, Hoffman BM. 1987. Yeast cytochrome *c* with phenylalanine or tyrosine at position 87 transfers electrons to (zinc cytochrome *c* peroxidase)⁺ at a rate ten thousand times that of serine-87 or glycine-87 variants. *Proceedings of the National Academy of Sciences of the United States of America* 84: 1249-1252.

Pielak GJ, Concar DW, Moore GR, Williams RJP. 1987. The structure of cytochrome *c* and its relation to recent studies of long-range electron transfer. *Protein Engineering* 1: 83-88.

Pielak GJ, Oikawa K, Mauk AG, Smith M, Kay CM. 1986. Elimination of the negative Soret Cotton effect of eukaryotic cytochromes *c* by replacement of an invariant phenylalanine residue by site-directed mutagenesis. *Journal of the American Chemical Society* 108: 2724-2727.

Pielak GJ, Gurusiddiwiah S, Legg JI. 1986. The decomposition of azo lysine. *Analytical Biochemistry* 156: 403-405.

Pielak GJ, Mauk AG, Smith M. 1985. Site-directed mutagenesis of cytochrome *c* shows that an invariant phe is not essential for function. *Nature* 313: 152-154.

Pielak GJ, Urdea MS, Igi K, Legg JI. 1984. Azo protein analogs: synthesis and characterization of arsanilazo and sulfanilazo derivatives of tyrosine and histidine. *Biochemistry* 23: 589-596.

Pielak GJ, Urdea MS, Legg JI. 1984. Preparation and characterization of arsanilazo and sulfanilazo proteins. *Biochemistry* 23: 596-603.

Legg JI, Igi K, Pielak GJ, Warner BD, Urdea MS. 1980. Circular dichroism as a probe of metal Ion interactions with azo proteins. In: Douglas BE, Yoshihiko S, eds. *Stereochemistry of Optically Active Transition Metal Compounds*. Washington DC: American Chemical Society *pp. 195-205*.